

Effects of antifungal activity of *Daenensis* thyme (*Thymus daenensis*) and *Mentha* (*Mentha longifolia*) essential oils on rainbow trout (*Oncorhynchus mykiss*) eggs hatchability

M Salehi¹, M Soltani² and S P Hosseini-Shekarabi¹

¹Department of Fisheries Science, Science and Research Branch, Islamic Azad University, Tehran, Iran.

²Department of Aquatic Animal Health, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran.

Received: April 2016

Accepted: September 2016

Abstract

In this study, rainbow trout (*Oncorhynchus mykiss*) eggs were treated with effective doses of *Daenensis* thyme, *Thymus daenensis* (20, 10 and 5 mgL⁻¹) and *Mentha*, *Mentha longifolia* (10, 5 and 2.5 mgL⁻¹), during incubation period (20 days) until hatching for 30 minutes per every other day as constant flow bath treatment method. The mortality level in normal control (without any treatment) was significantly higher than other groups ($p < 0.05$). Level of mortality in egg treated with *Daenensis* thyme at 20 mgL⁻¹ ($12.01 \pm 0.357\%$) was significantly lower than both normal control ($33.35 \pm 0.889\%$) and *Mentha* groups until the eyed-egg stage ($p < 0.05$). No significant different was seen between *Mentha* at 10 and 5 mgL⁻¹ mortality until eyed-egg stage ($p > 0.05$).

Correspondence: M Soltani, Department of Aquatic Animal Health, Faculty of veterinary medicine, University of Tehran, Tehran, Iran (e-mail: msoltani@ut.ac.ir)

The highest hatching rate ($78.36 \pm 0.340\%$) was recorded in *Daenensis* thyme at 20 mgL⁻¹ concentrations compared to other groups except Malachite green ($p < 0.05$). Eggs treated with *Daenensis* thyme and *Mentha* essence showed greater mean percentage of survival and hatching rates compared to the normal control ($p < 0.05$). However, essential oil derived from *Daenensis* thyme at 20 mgL⁻¹ probably has the potential to be used as health control of rainbow trout eggs against fungal contamination.

Keywords: Antifungal activity, *Daenensis* thyme, Essential oil, Malachite green, *Mentha*.

Introduction

One of the main types of fungal diseases in farmed salmonid fish is saprolegniasis and causes considerable economic loss in the fish farming, especially hatchery sector (Bruno & Wood 1999; Pottinger & Day 1999). The occurrence and severity of saprolegniasis depend on the water sources, water

temperature, organic load and length of contact time (Ahmadi, Hajimoradloo, Ghorbani, Chitsaz & Soleimani 2012). Malachite green has been used extensively by the aquaculture industry as an effective fungicide throughout the world for many years in the past (Pottinger & Day 1999). However, Malachite green is considered carcinogenic, mutagenic, teratogenic and harmful bioaccumulation chemical material (Meyer & Jorgenson 1983; Meinertz, Stehly, Gingerich & Allen 1995; Culp, Mellick, Trotter, Greenlees, Kodell & Beland 2006; Sudova 2007; Carral, Gonzalez, Celada, Saez-Royuela, Melendre, Gonzalez & Garcia 2009). The loss of this extremely effective substance in the fish farming industry has driven investigators to look for a non-hazardous material and environmentally friendly which is as effective as malachite green such as medical plants. *Thymus* species are well known as medicinal plants because of having biological and pharmacological properties (Stahl-Biskup & Saez 2002). Daenensis thyme, *Thymus daenensis*, is an endemic aromatic medicinal plant to Iran. Essential oil of *T. daenensis* is a rich source of thymol, which gives high antimicrobial and antioxidant activity to this plant (Akbarinia & Mirza 2008). Essential oil from *Mentha* (*Mentha longifolia*) is generally used in flavors and fragrances (Iskan, Kirimer, Kurkcuglu, Baser & Demirci 2002) and furthermore the antibacterial properties of the essential oil from the leaves were recorded (Cowan 1999). Several studies have concluded that plant essential oils have greater antimicrobial

activity than chemical components (Davidson & Parish 1989; Gill, Delaquis, Russo & Holley 2002). Moreover, a number of reports have been issued on the effect of antifungal activity of different herbal oils on aquatic eggs such as rainbow trout eggs (Sharif-Rohani, Ebrahimzadeh-Mousavi, Khosravi, Mokhayer, Bahonar, Mirzargar & Mehrabi 2006; Mousavi, Mirzargar, Ebrahimzadeh-Mousavi, Omid Baigi, Khosravi, Bahonar & Ahmadi 2009; Khosravi, Shokri, Sharifrohani, Ebrahimzadeh-Mousavi & Mousavi 2012), narrow-clawed crayfish (Koca & Cevikbas 2014), Persian Sturgeon (Ahmadi et al. 2012), Iberian rock lizard (Moreira & Barata 2005) and Kutum (*Rutilus frisii kutum*) (Najafi & Zamini 2013).

Nowadays, world follow using safe, effective and eco-friendly substances with decreasing of chemical disinfections. For instance, Khosravi et al. (2012) reported that *Zataria* (*Zataria multiflora*) essential oil (5 and 10 mgL⁻¹) and *Eucalyptus camaldolensis* (25 mgL⁻¹) increased hatching rates on rainbow trout eggs infected with *Saprolegnia parasitica*. Mousavi et al. (2009) indicated inhibitory effects of the combination of essential oils (*Thymus vulgaris*, *Salvia officinalis*, *Eucalyptus globulus* and *Mentha piperita*) on the filamentous fungi and increase hatching rate in comparison with malachite green on rainbow trout eggs on concentration 10 mgL⁻¹. Moreover, there is a report indicating that *Zataria* essential oil has immunostimulatory effects in common carps (Soltani, Sheikhzadeh, Ebrahimzadeh-Mousavi & Zaegar 2010).

Chemical disinfections are routinely used in Iranian rainbow trout reproduction centers for controlling of the eggs fungal contamination. This study was intended to assess, two different herbal essential oils, including *Daenensis* thyme (*T. daenensis*) and *Mentha* (*M. longifolia*), as alternative options against saprolegniasis on rainbow trout (*Oncorhynchus mykiss*) eggs.

Materials and Methods

This experiment was carried out at the Abzi Exir Kowsar propagation and breeding center (Lorestan Province, Iran). Broodstock rainbow trout were anesthetized with clove powder (120 mgL^{-1}) and eggs from females ($1.6 \pm 0.5 \text{ kg}$ average weight) were stripped into a dry bowl and fertilized with milt from a ripe male ($1 \pm 0.2 \text{ kg}$ average weight). The mean water temperature was maintained within $13 \pm 0.5^\circ\text{C}$, dissolved oxygen concentration, pH and ammonia nitrogen (AN) were calculated approximately $8 \pm 1 \text{ mgL}^{-1}$, 7.8- 8.1, less than 0.01 mgL^{-1} during the experiment, respectively. Temperature, dissolved oxygen and pH were measured using by a portable multi meter (Instrument Corp. 8603, Taiwan). The water flow rate was adjusted at 0.8 liter per minute with aeration provided throughout the incubation and hatching period.

The *T. vulgaris* and *M. longifolia* oils obtained with hydrodistillation method for 4 hours using a Clevenger-type apparatus to produce essential oils according to Pirbalouti, Malekpoor, Enteshari, Yousefi, Momtaz & Hamed (2010). Phenytoin sodium (20 mgkg^{-1})

was added to the essential oils before using to dissolve in water (Mousavi et al. 2009; Soltani et al. 2010).

All incubators were completely cleaned and disinfected with formalin, before transferring eggs into incubators. The fertilized eggs (8.4 kg) were then divided to 24 groups, randomly. The treatment trials started 24 hours post-incubation and were continued until hatching stage. Three concentrations of *Daenensis* thyme ($5, 10, 15 \text{ mgL}^{-1}$) and *Mentha* ($2.5, 5, 10 \text{ mgL}^{-1}$) were used in this study against mold contamination. All treatments were applied for 30 minutes per every other day as constant flow treatment method. Malachite green (Merck, Germany) was used as a positive control at concentration of 2 mgL^{-1} (Kitancharoen, Yamamoto & Hatai 1998). There was no treatment in normal control group. The experiment was continued for 20 days (from fertilized eggs to hatching stage). During the experiment, no handling and transferring of eggs were performed (Mousavi et al. 2009).

During this period, eggs were monitored everyday and the ones showing mold contamination were counted. At the end of incubation period, hatching rate of eggs was calculated.

All data were subjected to a one-way analysis of variance (ANOVA). Significance of the differences between means was tested using Duncan's multiple range test ($p < 0.05$). Each treatment was three replicate and sample volume in each stage of the treatment was 40%.

Results

Effects of different disinfectant materials on the successful of hatching during different stages are shown in Table 1. Among essential oils treatments, *Daenensis* thyme at 20 mgL⁻¹ (12.01±0.357%) showed the lowest mortality among eggs until eyed-egg stage (p<0.05). But the lowest mortality was observed in Malachite green group (11.29±0.639%), which it was significant than the other treatments (p<0.05). The lowest mortality of eggs from eyed-egg to hatching stage was belong Malachite green (6.93±0.354%), followed by *Daenensis* thyme at 20 mgL⁻¹

(9.24±0.299%)(p<0.05).In contrast, the highest mortality was obtained in normal control treatment in all egg stages (Table 1; p<0.05).

In surveying of hatching rate, the highest were Malachite green and *Daenensis* thyme at 20 mgL⁻¹ treatments with 81.78±0.827 % and 78.36±0.340 %, respectively. However, the lowest ratio was significantly observed in control (51.61±0.153%) than other treatments (Fig. 1; p<0.05).

All the essential oils demonstrated fungi static properties compared to normal control (p<0.05).

Table 1 Hatching success of rainbow trout eggs among different treatments based on different concentrations of essential oils and Malachite Green (Mean±SD)

Group	Concentration (mgL ⁻¹)	Initial eggs (No.)	Total mortality of eggs (No.)	Mortality of eggs until eyed-egg stage (%)	Mortality of eggs from eyed-egg to hatching stage (%)
<i>Daenensis</i> thyme	20	4406±9 ^{ab}	936±4 ^b	12.01±0.357 ^b	9.24±0.299 ^b
	10	4417±21 ^c	1016±7 ^{bc}	12.93±0.447 ^{bc}	10.08±0.547 ^{bc}
	5	4404±30 ^a	1032±18 ^c	13.08±0.098 ^c	10.36±0.376 ^{bd}
<i>Mentha</i>	10	4417±18 ^c	1299±2 ^{cd}	20.04±0.144 ^d	9.37±0.256 ^c
	5	4422±14 ^{ab}	1369±20 ^d	20.10±0.286 ^d	10.47±0.229 ^d
	2.5	4433±18 ^d	1391±20 ^d	20.77±0.576 ^{dc}	10.62±0.261 ^d
Malachite green	2	4408±19 ^b	803±34 ^a	11.29±0.639 ^a	6.93±0.354 ^a
Normal control	-	4428±9 ^{ba}	2143±16 ^f	33.35±0.889 ^f	15.04±0.603 ^f

Values with differing letters are significantly different (p<0.05) from the other values in the same column.

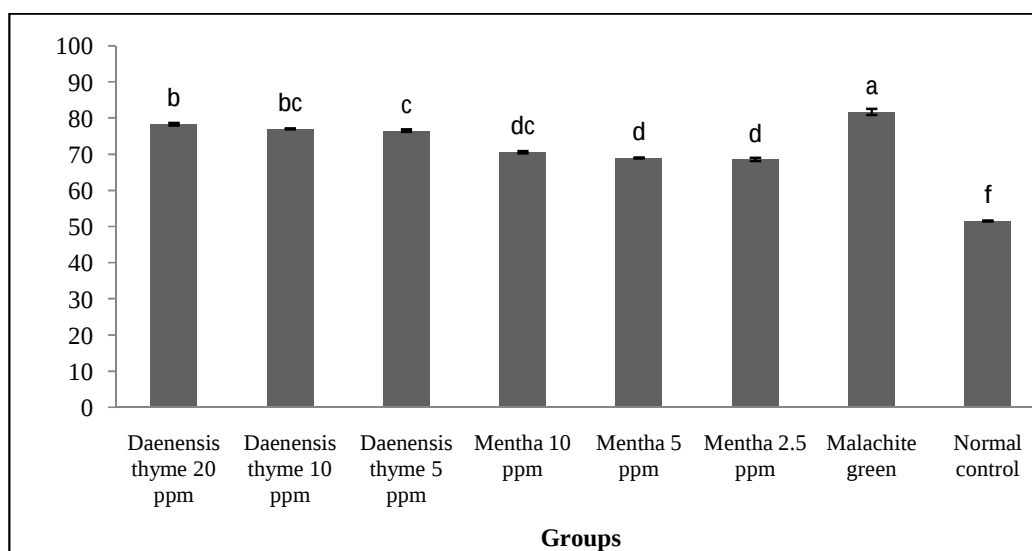


Figure 1 Effects of essential oils on hatching rate (%) of rainbow trout eggs at 20 days (Mean±SD). Different upper case superscripts indicate significant differences ($p < 0.05$).

Discussion

Fungal infestations can be controlled chemically with antifungal agents and physically by removing fluffy, cotton-like, and white to grayish dead eggs. Manual removal of dead eggs can be safely performed only after the eyed stage (Jensen & Alderdice 1989) and therefore the spreading of fungi before this period must be prevented by other means. In modern large-scale fish hatcheries, chemical control of fungi is the preferred method (Hoskonen, Heikkinen, Eskelinen & Pirhonen 2013). Several investigators concluded that essential oils have greater antimicrobial (Davidson & Parish 1989; Gill et al. 2002) and aquatic fungicidal (Mori, Hirose, Hanjavanit & Hatai 2002; Sharif-Rohani et al. 2006; Khosravi et al. 2008; Mousavi et al. 2009; Ahmadi et al. 2012; Khosravi et al. 2012) activities than some chemical components.

Malachite green is used as the best chemical material in controlling of rainbow trout eggs mold infection for many years, however, Meinertz et al. (1995) concluded that undetectable residues of malachite green would still remain in fish grown from eggs which had been exposed to the chemical until they reached market size. Moreover, Andersen, Roybal & Turnipseed (2005) found residues of malachite green and of its conversion product leucomalachite green in market size salmon, and because of concerns regarding the consumers and the health implications of the operators at the fish farms the use of it was banned by FDA since 1991. Therefore, there has been significant effort expended to identify natural therapeutic agents, being effective as malachite green.

Data analysis showed that the best hatching rate among essential oils experiments was recorded in *Daenensis thyme* at 20 mgL⁻¹ when

the rainbow trout eggs were bathed for 30 min. Based on previous report (Moreira & Barata 2005), natural monoterpene phenol like thymol is the main components of *Daenensis* thyme essential oil and it has been associated with strong antimicrobial attributes (Palaniappan & Holley 2010). The possible mechanisms of phenolic compounds are appeared to bedegradation of the cell wall, damage to cytoplasmic membrane, damage to membrane proteins, leakage of cell contents and depletion of the protein motive force (Burt 2004).

In similar studies, Mousavi et al. (2009) reported that Eucalyptus essential oilin dose of 200 mgL⁻¹ is effective in control of Saprolegniasis and resulted in higher hatching rate in rainbow trout eggs. Najafi & Zamini (2013) reported that dosage 100 mgL⁻¹ Eucalyptus essential oil controlled *Rutilus frisii kutum* eggs infected with Saprolegnia. In the other experiment, Eucalyptus essential oil increased hatchability of *Acipenser persicus* eggs with concentration of 200 mgL⁻¹ (Ahmadi et al. 2012). Bouchard, Patel & Lahey (2000) found that rainbow trout eyed eggs treated with clove oil at 1 g/lit had the lowest rate of fungus infection. Similarly, Khosravi et al. (2012) showed thart *Zataria multiflora* at concentration of 25 mgL⁻¹, *E. camaldolensis* at concentration of 25 mgL⁻¹, and *Geranium herbarium* at concentration of 100mgL⁻¹ for 60 min daily were the best treatments to the prevention of fungal attack, the increase of hatching rate and the eyed egg survival rate of rainbow trout egg. However, the recommended doses of herbal essential oils

used in the above studies to improve hatchability of fish eggs are significantly higher than the doses obtained from *Daenensis* thyme in the present study. This result illustrated that antibacterial and antifungal actives of *Daenensis* thyme may higher than Eucalyptus, clove, *Zataria* and *Geranium* essential oils according to its specific components. In contrast, *Satureja cuneifolia* essential oil showed highest hatching rates (44.5±0.57%) in infected rainbow trout egg with *Saprolegnia parasitica* in lower concentration (5 mgL⁻¹) (Metin, Diler, Dıdinen, Terzioğlu & Gormez 2013).

According to the results, all essential oils especially *Daenensis* thyme group improved mortality rate of the eggs until eyed-egg stage compared to normal control. This finding recommended that using of *Daenensis* thyme essential oil after eyed-egg stage is safe and inhibit spreading of fungi after this period. In contrast, some chemical agents such as hydrogen peroxide treatments during blastopore formation increase rainbow trout egg mortality making the chemical better suited to use after the eyed-egg stage (Gaikowski, Rach, Olson, Ramsay & Wolgamood et al. 1998) or the concentration should be decreased during estimated blastopore formation (Barnes & Gaikowski 2003).

The results showed that treatments of rainbow trout eggs with Malachite green (2 mgL⁻¹) greatly improved the hatching success compared to the essential oils groups. This finding is in agreement with other studies (Sharif-Rohani et al. 2006; Mousavi et al. 2009; Khosravi et al. 2012; Najafi & Zamini

2013) which are reported Malachite green at 2 mgL⁻¹ treatment gave the best hatching performance in comparison to essential oils treatments. However, toxicology and teratology effects of malachite green on fish and other animals have been reported (Andersen et al. 2005; Culp et al. 2006; Sudova 2007). However, there is not any report for toxicity effect of the herbal extract and essential oil on fish and human up to present. Therefore, the essential oils can be a potential substitute for controlling filamentous fungi unlike chemical agents in aquaculture. The Daenensis thyme essence with 20 mgL⁻¹ can decrease mold infection rate and increase hatch rates in hatcheries and may represent alternative therapeutic treatments in rainbow trout aquaculture and hatchery sectors. Essential oils of this study need more trials for evaluating toxicological effects on rainbow trout eggs at high doses.

References

- Ahmadi M., Hajimoradloo A., Ghorbani R., Chitsaz H. & Soleimani H. (2012) Effects of Eucalyptus Essence, Malachite Green and Sodium Chloride on Hatching Rate of Persian Sturgeon (*Acipenser persicus*) Eggs. *Journal of Veterinary Advances* 10, 488-493.
- Akbarinia A. & Mirza M. (2008) Identification of essential oil components of *Thymus daenensis* Celak. In field condition in Qazvin. *The Journal of Qazvin University of Medical Sciences* 3, 58-62.
- Andersen W.C., Roybal J.E. & Turnipseed S.B. (2005) Liquid Chromatographic Determination of Malachite Green and Leucomalachite Green (LMG) Residues in Salmon with in situ LMG Oxidation. *Journal of AOAC International* 88, 1292-1298.
- Barnes M.E. & Gaikowski M.P. (2003) Use of hydrogen peroxide during incubation of landlocked fall chinook salmon eggs in vertical-flow incubators. *North American Journal of Aquaculture* 66, 29-34.
- Boyd C.E. & Tucker C.S. (1992) *Water Quality and Pond Soil Analysis for Aquaculture*. Alabama Agricultural Experiment Station, Auburn University Publications, USA.
- Bouchard L., Patel, J. & Lahey, L. (2000) The effect of clove oil on fungal infections of salmonid eggs. *Bulletin of the Aquaculture Association of Canada* 4, 110-112.
- Bruno D.W. & Wood B.P. (1999) *Saprolegnia and other Oomycetes*. In: Fish diseases and disorders viral, bacterial and fungal infections (ed. by P. T. K. Woo & D. W. Bruno), pp. 599-659. CABI Publishing, Wallingford, Owon, United Kingdom.
- Burt S. (2004) Essential oils: Their antibacterial properties and potential applications in foods: A review. *International Journal of Food Microbiology* 94, 223-253.
- Carral J. M., Gonzalez A., Celada J.D., Saez-Royuela M., Melendre P.M., Gonzalez R. & Garcia V. (2009) Antifungal treatments in

- artificial incubation of crayfish eggs (*Pacifastacus leniusculus*, Astacidae): searching for alternatives to formalin. *Knowledge and Management of Aquatic Ecosystems* 16, 394-395.
- Cowan, M. M. (1999) Plant Products As Antimicrobial Agents. *Clinical Microbiology Reviews* 12, 564-582.
- Culp S. J., Mellick P. W., Trotter R. W., Greenlees K. J., Kodell R. L. & Beland F. A. (2006) Carcinogenicity of malachite green chloride and leucomalachite greenin B6C3F1 mice and F344 rats. *Food and Chemical Toxicology* 44, 1204-1212.
- Davidson P. M. & Parish M. E. (1989) Methods for testing the efficacy of food antimicrobials. *Food Technology* 43, 148-155.
- Gaikowski M. P., Rach J. J., Olson J. J., Ramsay R. T. & Wolgamood M. (1998) Toxicity of hydrogen peroxide treatments to rainbow trout eggs. *Journal of Aquatic Animal Health* 3, 241-251.
- Gill A. O., Delaquis P., Russo P. & Holley R. A. (2002) Evaluation of antilesterial action of cilantro oil on vacuum packed ham. *International Journal of Food Microbiology* 73, 83-92.
- Hoskonen P., Heikkinen J., Eskelinen P. & Pirhonen J. (2013) Efficacy of clove oil and ethanol against *Saprolegnia* sp. and usability as antifungal agents during incubation of rainbow trout *Oncorhynchus mykiss* (Walbaum) eggs. *Aquaculture Research* 46, 581-589.
- Iscan G., Kirimer N., Kurkcuoglu M., Baser K. H. & Demirci F. (2002) Antimicrobial screening of *Menthapiperita* essential oils. *Journal of Agricultural and Food Chemistry* 50, 3943-3946.
- Khosravi A. R., Eslami A.R., Shokri H. & Kashanian M. (2008) *Zataria multiflora* cream for the treatment of acute vaginal candidiasis. *International Journal of Gynecology and Obstetrics* 101, 201-202.
- Khosravi A. R., Shokri H., Sharifrohani M., Ebrahimzadeh-Mousavi H. E. & Mousavi Z. (2012) Evaluation of the antifungal activity of *Zataria multiflora*, *Geranium herbarium*, and *Eucalyptus camaldolensis* essential oils on *Saprolegnia parasitica*–infected rainbow trout (*Oncorhynchus mykiss*) eggs. *Foodborne Pathogens and Disease* 7, 674-679.
- Kitancharoen N., Yamamoto A. & Hatai K. (1998) Effects of sodium chloride, hydrogen peroxide and malachite green on fungal infection in rainbow trout eggs. *Biocontrol Science and Technology* 3, 113-115.
- Koca S. B. & Cevikbas M. (2014) Antifungal effect of *Origanum onites* essential oil as an alternative to formalin in the artificial incubation of narrow-clawed crayfish (*Astacus leptodactylus* Eschscholtz, 1823). *Aquaculture Research* 46, 2204-2210.
- Meinertz J.R., Stehly G.R., Gingerich W.H. & Allen J.L. (1995) Residues of malachite green

in eggs and fry of rainbow trout, *Oncorhynchus mykiss* (Walbaum), after treatment of eggs. *Journal of Fish Diseases* 18, 239-247.

Metin S., Diler O., Didinen B. I., Terzioğlu S. & Gormez O. (2013) In vitro and in vivo antifungal activity of *Satureja cuneifolia* ten essential oil on *Saprolegnia parasitica* strains isolated from rainbow trout (*Oncorhynchus mykiss*, Walbaum) eggs. *Aquaculture Research* 46, 1-7.

Meyer F.P. & Jorgenson T.A. (1983) Teratological and other Effects of Malachite Green on Development of Rainbow Trout and Rabbits. *Transactions of the American Fisheries Society* 112, 818-824.

Moreira P.L. & Barata M. (2005) Egg mortality and early embryo hatching caused by fungal infection of Iberian rock lizard (*Lacerta monticola*) clutches. *Herpetological Journal* 15, 265-272.

Mori T., Hirose H., Hanjavanit C. & Hatai K. (2002) Antifungal activities of plant extracts against some aquatic fungi. *Biocontrol Science and Technology* 7, 187-191.

Mousavi S.M., Mirzargar S.S., Ebrahimzadeh-Mousavi H., Omid Baigi R., Khosravi A., Bahonar A. & Ahmadi M.R. (2009) Evaluation of Antifungal Activity of New Combined Essential Oils in Comparison With Malachite Green on Hatching Rate in Rainbow Trout (*Oncorhynchus mykiss*) Eggs. *Journal of fishery and aquatic science* 2, 103-110.

Najafi M. & Zamini A.A. (2013) Comparative analysis of antifungal properties of *Zataria multiflora*, Eucalyptus spp Essence and Malachite Green on Eggs of Kutum (*Rutilus frisii* Kutum). *Advances in Biological Research* 5, 163-168.

Palaniappan K. & Holley R.A. (2010) Use of natural antimicrobials to increase antibiotic susceptibility of drug resistant bacteria. *International journal of food microbiology* 140, 164-168.

Pirbalouti A.G., Malekpoor F., Enteshari S., Yousefi M., Momtaz H. & Hamed, B. (2010) Antibacterial activity of some folklore medicinal plants used by Bakhtiari tribal in Southwest Iran. *International Journal of Biology* 2, 55-63.

Pottinger T.G. and Day J.G. (1999) A *Saprolegnia parasitica* challenge system for rainbow trout: assessment of Pyceze as an anti-fungal control agent for both fish and ova. *Diseases of Aquatic Organisms* 36, 129-141.

Sharif-Rohani M., Ebrahimzadeh Mousavi H., Khosravi A., Mokhayer B., Bahonar A., Mirzargar S. & Mehrabi Y. (2006) Evaluation of *Geranium herbarum* essence application in control of fungal contamination in Rainbow trout (*Oncorhynchus mykiss*) eggs. *Journal of Faculty of Veterinary Medicine* 61, 269- 272.

Soltani M., Sheikhzadeh N., Ebrahimzadeh-Mousavi H.A. & Zaegar A. (2010) Effect of *Zataria multiflora* Essential oil on innate immune responses of common carp (*Cyprinus carpio*).

Journal of Fisheries and Aquatic Science 5, 191-199.

Stahl-Biskup E. & Saez F. (2002) Thyme: The genus *Thymus*. CRC Press, New York, 354.

Sudova E., Machova J., Svobodova Z. & Vesely T. (2007) Negative effects of malachite green and possibilities of its

replacement in the treatment of fish eggs and fish: a review. *Veterinarni Medicina* 52, 527-539.

Wolgamood M. (1998) Toxicity of hydrogen peroxide treatments to rainbow trout eggs. *Journal of Aquatic Animal Health* 10, 241-251.

اثر ضدقارچی اسانس های آویشن دنایی (*Thymus daenensis*) و پونه کوهی (*Mentha longifolia*) در قابلیت تخم گشایی ماهی قزل آلاي رنگين کمان

میثم صالحی¹، مهدی سلطانی^{2*} و سید پژمان حسینی شکرابی¹

1-دانشگاه آزاد اسلامی، واحد علوم و تحقیقات تهران، گروه شیلات، تهران، ایران.
2-گروه بهداشت و بیماریهای آبزیان، دانشکده دامپزشکی، دانشگاه تهران، تهران، ایران

چکیده:

دراین مطالعه، تخم ماهی قزل آلاي رنگين کمان با غلظت های موثر از اسانس های آویشن دنایی (20، 10 و 5 میلی لیتر در لیتر) و پونه کوهی (10، 5 و 2/5 میلی لیتر در لیتر) درطول دوره انکوباسیون تا زمان تخمگشایی (20 روز) به مدت 30 دقیقه به صورت حمام یکرز در میان با آب جریاندار تحت درمان قرارگرفتند. میزان مرگ و میر (بدون تداخل دارویی) به طورمعنی داری اذتمام تیمارها بیشتر بود ($p<0/05$). میزان مرگ و میر تخم در تیمار درمان شده با اسانس آویشن دنایی با غلظت 20 میلی لیتر در لیتر (12/01%) به طور معنی داری از تیمار کنترل ($33/35\pm0/889\%$) و تیمارهای پونه کوهی تا مرحله چشم زدگی تخم کمتر بود ($p<0/05$). اختلاف معنی داری بین میزان مرگ و میر تیمارهای پونه کوهی با غلظت 10 و 5 میلی لیتر در لیتر تا مرحله چشم زدگی مشاهده نشد ($p<0/05$). بیشترین نرخ تخم گشایی در آویشن دنایی با غلظت 20 میلی لیتر در لیتر درمقایسه با سایر تیمارها بجز تیمار سبز مالاشیت ثبت شد ($p<0/05$). تخم های درمان شده با اسانس های آویشن دنایی و پونه کوهی میانگین درصدبقا و تخمگ شایی بالاتری نسبت به تیمار کنترل نشان دادند ($p<0/05$). در حالی که، احتمالاً اسانس آویشن دنایی باغلظت 20 میلی لیتر در لیتر پتانسیل استفاده شدن به عنوان یک ماده کنترل کننده سلامت تخم ماهیان قزل آلاي رنگين کمان درمقابل آلودگی های قارچی را دارد.

کلمات کلیدی: فعالیت ضدقارچی، آویشن دنایی، اسانس، سبز مالاشیت، پونه کوهی.

* نویسنده مسئول: msoltani@ut.ac.ir